

## Evaluation of the Impact of the Biofield Energy Treated Test Formulation on Various Biomarkers in human Bones, Heart, Liver, Lungs, and Brain Cells

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**Keywords** The Trivedi effect<sup>®</sup>, Biofield energy treatment; Cardiac health; Liver health; Lungs health; VDR receptor; Brain health; Bone health

## Abstract

Vital organs dysfunctions are the major concern for human health worldwide. The study aim was to investigate the impact of Biofield Treated test formulation on vital organs function using cell-based assays. The test formulation/test item (TI) and cell media (Med) was divided into two parts; one untreated (UT) and other part received the Biofield Treatment remotely by a renowned Biofield Energy Healer, Victoria Lee Vannes, USA and was labeled as Biofield Treated (BT) test formulation/media. Based on the cell viability test formulation was found safe in six different cells. The test formulation groups showed 112.6% and 108.65% restoration of cell viability in human cardiac fibroblasts cells (HCF); while, 845.63% restoration of cell viability in human hepatoma cells (HepG2) compared to UT-Med + UT-TI group. Furthermore, 131.86% restoration of cell viability was observed in adenocarcinomic human alveolar basal epithelial cells (A549) than untreated. The alkaline phosphatase (ALP) level was significantly increased by 94.87%, 99.06%, and 105.13% in UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively at 10 µg/mL in human bone osteosarcoma cells (MG-63) than untreated. Additionally, ALP level was significantly increased by 150.97%, 382.08%, and 471.4% in UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI, respectively at 0.1 µg/mL in human endometrial adenocarcinoma cells (Ishikawa) than untreated. The percent protection of HCF (heart) cells (decreased of LDH activity) was significantly increased by 115.1% (1 µg/mL) and 165.77% (10 µg/mL) in BT-Med + UT-TI and BT-Med + BT-TI, respectively than untreated.

The percent protection of HepG2 (liver) cells (decreased of ALT activity) was significantly increased by 117.65% (1 µg/mL) and 91.3% (63 µg/mL) in UT-Med + BT-TI and BT-Med + BT-TI, respectively than untreated. The percent protection of A549 (lungs) cells (increased of SOD activity) was significantly increased by 40.56% in UT-Med + BT-TI at 10 µg/mL than untreated. Serotonin level was significantly increased by 543.84% (1 µg/mL), 477.12% (10 µg/mL), and 457.22% (10 µg/mL) in UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI, respectively than untreated. The relative quantification (RQ) of vitamin D receptor (VDR) was significantly increased by 186.96%, 341.43%, and 291.31% in UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI, respectively at 1 µg/mL than untreated. Overall, these results suggest that Biofield Treated test formulation significantly improved the bones, heart, liver, lungs, and brain-related functional enzyme biomarkers. Therefore, the Trivedi Effect<sup>®</sup> can be used as a complementary and alternative therapy against several disorders such as coronary artery disease, heart attack, arrhythmias, heart failure, congenital heart disease, cardiomyopathy, cirrhosis, liver cancer, hemochromatosis, Wilson disease, pneumonia, asthma, emphysema, chronic bronchitis, cystic fibrosis, osteoporosis, etc.

## Introduction

Bones, heart, liver, lungs, and brain disorders are the major concern of human overall health across the globe. The World Health Organization (WHO) estimates, in 2016, ~17.5 million people die due to cardiovascular (heart) disorders, ~3.5 million people die due to lungs disorders, ~1.3 million people die due to liver disorders around the globe each year [1]. Moreover, ~1.2 million people most frequently diagnosed adult-onset brain disorders in each year in the USA. [2]. Three main criteria to keep a healthy heart include the opening blood vessels, strengthening the heart muscle, and controlling free radical damage by antioxidants [3]. The release of liver mitochondrial enzymes is considered strong evidence for hepatic (liver) necrosis, which is associated with an increased production of reactive oxygen species (ROS) that leads to hepatic lipid peroxidation [4-6]. Oxidative stress in the respiratory system increases the production of mediators of pulmonary inflammation and initiate or promote mechanisms of carcinogenesis [7]. The lung is one of the major organs, which is highly exposed by various oxidants i.e., endogenous and exogenous oxidants (cigarette smoke, mineral dust, ozone, and radiation). These oxidants produce free radicals, while reactive oxygen species (ROS) and reactive nitrogen species (RNS) are produced by phagocytes as well as by alveolar, polymorphonuclear, bronchial and different endothelial cells [8]. However, the role of oxidative stress in the pathogenesis of lung diseases has been widely reported such as asthma,

chronic obstructive pulmonary disease (COPD), lung malignancies and parenchymal lung diseases like idiopathic pulmonary fibrosis and lung granulomatous diseases [9]. Serotonin (5-hydroxytryptamine, 5-HT) is among the brain's neuromodulators responsible for behavior and understanding [10]. Apart from medicines, non-pharmacologic methods can increase serotonin by increasing recognition and happiness and well-being. These factors can protect against mental and physical disorders [11]. There is currently no universally accepted test formulation, which improve the organ health biomarkers. With this respect, the novel test formulation was designed on the basis of best scientific literature, which is the combination of herbal products *viz.* panax ginseng extract and beta carotene, minerals *viz.* calcium chloride, magnesium gluconate, zinc chloride, sodium selenate, ferrous sulfate, and vitamins *viz.* vitamin B12, vitamin D3, ascorbic acid, and vitamin B6. This formulation is designed for overall functioning of the organs that can results in improved overall health conditions against many pathological conditions such as lung disorder, liver disorder, breast cancer, liver cancer, aging, muscle damage, and overall health. Minerals and vitamins present in the test formulation provide significant functional support to all the vital organs [12-14]. In addition, panax ginseng is one of the best reported medicinal plants that improve mental, physical abilities, cognitive health, and is potent immunomodulator [15,16].

Various study data suggested the effect of Energy Therapy in cancer patients through therapeutic touch [17], massage therapy [18], etc. Complementary and Alternative Medicine (CAM) therapies are preferred model of treatment, among which Biofield Therapy (or Healing Modalities) is one approach to enhance emotional, mental, physical, and human wellness. The National Center of Complementary and Integrative Health (NCCIH) has recognized and allowed Biofield Energy Healing as a CAM approach in addition to other therapies and medicines such as natural products, chiropractic/osteopathic manipulation, Qi Gong, deep breathing, Tai Chi, yoga, meditation, massage, special diets, healing touch, relaxation techniques, traditional Chinese herbs and medicines, naturopathy, movement therapy, homeopathy, progressive relaxation, guided imagery, pilates, acupuncture, acupressure, Reiki, rolfing structural integration, hypnotherapy, Ayurvedic medicine, mindfulness, essential oils, aromatherapy, and cranial sacral therapy. The Human Biofield Energy has subtle energy that has the capacity to work in an effective manner [19]. CAM therapies have been practiced worldwide with reported clinical benefits in different health disease profiles [20]. This energy can be harnessed and transmitted by the practitioners into living and non-living things via the process of Biofield Energy Healing. The Biofield Energy Treatment, the Trivedi Effect®, has been reported to have a significant impact in the field of cancer research [21,22], materials science [23, 24], microbiology [25,26], agriculture [27,28], nutraceuticals [29,30], and biotechnology [31,32]. Further, the Trivedi Effect® also significantly improved bioavailability of various low bioavailable compounds [33-35], an improved overall skin health [36,37], bone health [38-40], human health and wellness. Based on the excellent outcomes of the Biofield Energy Therapy in wide spectrum of areas, the authors intend to see the impact of the Biofield Energy Healing Treated test formulation on the function of vital organs such as bones, heart, liver, lungs, and brain specific biomarkers in different cell-lines.

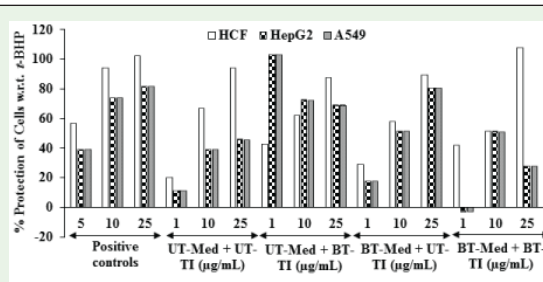
## Materials and Methods

### Chemicals and reagents

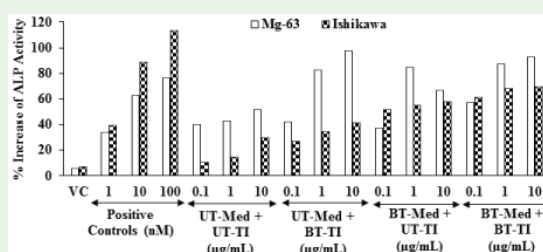
Ferrous sulfate, vitamin B6, vitamin D3, vitamin B12, calcium chloride, naringenin, trimetazidine (TMZ), 3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide (MTT), and ethylenediaminetetraacetic acid (EDTA) were obtained from Sigma Chemical Co. (St. Louis, MO). Zinc chloride, magnesium gluconate,  $\beta$ -carotene, and calcitriol were purchased from TCI chemicals, Japan. Panax ginseng extract obtained from panacea Phytoextracts, India. Sodium selenate and ascorbic acid were obtained from Alfa Aesar, India. Silymarin and curcumin were obtained from Sanat Chemicals, India and quercetin obtained from Clearsynth, India. Reverse Transcription Kit, RNeasy Mini Kit, and Syber Green PCR kits were procured from Quagen, India. All the other chemicals used in this experiment were analytical grade procured from India.

### Biofield energy healing strategy

The test formulation was the combination of eleven ingredients *viz.* calcium chloride, panax ginseng extract, vitamin B12,  $\beta$ -carotene, vitamin D3, zinc chloride, magnesium gluconate, sodium selenate, ferrous sulfate, ascorbic acid, and vitamin B6. The test formulation and the cell media was divided into two parts; one untreated (UT) and other part received the Biofield Energy Treatment remotely by a



**Figure 1:** Assessment of cytoprotective effect of the test formulation in human cardiac fibroblasts cells (HCF), human hepatoma cells (HepG2), and adenocarcinomic human alveolar basal epithelial cells (A549) against *tert*-butyl hydroperoxide (*t*-BHP) induced damage. TMZ: Trimetazidine ( $\mu$ M), silymarin ( $\mu$ g/mL), and quercetin ( $\mu$ M) were used as positive control in HCF, HepG2, and A549 cells, respectively. UT: Untreated; Med: Medium; BT: Biofield Treated; TI: Test item.



**Figure 2:** The effect of the test formulation on alkaline phosphatase (ALP) in human bone osteosarcoma cells (MG-63) and human endometrial adenocarcinoma cells (Ishikawa). Calcitriol and naringenin were used as positive control in Mg-63 and Ishikawa cells, respectively. VC: Vehicle control; UT: Untreated; Med: Medium; BT: Biofield Treated; TI: Test item.

**Table 1:** Information related to six cell lines with their plating density and time-point.

S. No.	Cell Line	Plating	Time Point
1	MG-63 (Bone)	3x10 <sup>4</sup> cells/ well, 96-well plate	5 days
2	Ishikawa (Uterus)	3x10 <sup>4</sup> cells/ well, 96-well plate	5 days
3	A549 (Lung)	10x10 <sup>4</sup> cells/ well, 96-well plate	24 hours
4	HepG2 (Liver)	1x10 <sup>4</sup> cells/ well, 96-well plate	24 hours
5	Human Cardiac fibroblasts (Heart)	1x10 <sup>4</sup> cells/ well, 96-well plate	24 hours
6	SH-SY5Y (Neuronal cell)	10x10 <sup>4</sup> cells/ well, 96-well plate	24 hours

renowned Biofield Energy Healer, Victoria Lee Vannes, USA under laboratory conditions for ~3 minutes through healer’s unique Biofield Energy Transmission process and was labeled as the Biofield Energy Treated (BT) test formulation/media. Further, the untreated group was treated with a “sham” healer for comparison purposes. The “sham” healer did not have any knowledge about the Biofield Energy Healing Treatment. The Biofield Energy Healer was located in the USA, however the test items were located in the research laboratory of Dabur Research Foundation, New Delhi, India. Biofield Energy Healer in this experiment did not visit the laboratory, nor had any contact with the test samples. After that, the Biofield Energy Treated and untreated test items were kept in similar sealed conditions and used for the study as per the study plan.

**Assessment of cell viability using MTT assay**

Cells were counted using hemocytometer and plated in 96-well plates at the specific density described in Table 1. The cells were then incubated overnight under growth conditions to allow cell recovery and exponential growth. Following overnight incubation, cells were treated with different concentrations of test formulations (BT/ UT). Following respective treatments, cells were incubated in a CO<sub>2</sub> incubator at 37°C, 5% CO<sub>2</sub>, and 95% humidity and incubated for time period mentioned in Table 1. After incubation, the plates were taken out and 20 µL of 5 mg/mL of MTT solution was added to all the wells followed by additional incubation for 3 hours at 37°C. The supernatant was aspirated and 150 µL of DMSO was added to each well to dissolve formazan crystals. The absorbance of each well was read at 540 nm using Synergy HT microplate reader. The percentage cytotoxicity at each tested concentration of TI was calculated using Equation 1:

$$\% \text{ Cytotoxicity} = [(R-X)/R] * 100 \tag{1}$$

Where, X = Absorbance of treated cells; R = Absorbance of untreated cells

The concentrations exhibiting percentage cytotoxicity < 30% were considered as non-cytotoxic [41].

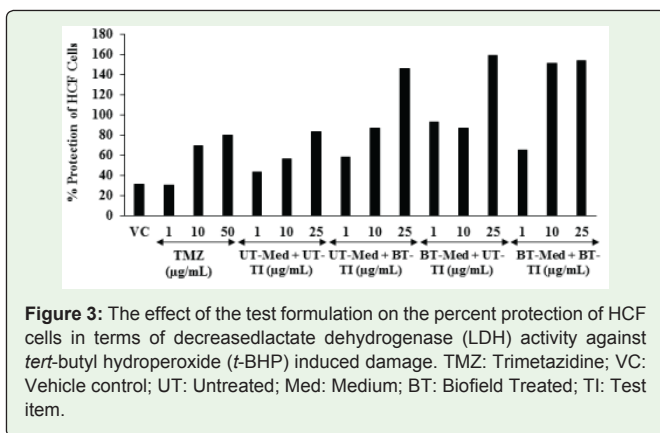
**Evaluation of the cytoprotective effect of the formulation**

Cells (human cardiac fibroblasts-HCF; human hepatoma cells-HepG2; and adenocarcinomic human alveolar basal epithelial cells-A549) were counted and plated in suitable medium followed by overnight incubation. The cells were then treated with the test items/ positive control at the non-cytotoxic concentrations for 24 hours. After 24 hours, oxidative stress was given to the cells using 10mM *t*-BHP for 3.5 hours. The untreated cells served as a control that did not receive any treatment and was maintained in cell growth medium only. Cells treated with 10mM of *t*-BHP alone served as negative control. After 3.5 hours of incubation with *t*-BHP the above plates were taken out and cell viability was determined by MTT assay. The percentage protection corresponding to each treatment was calculated using Equation 2:

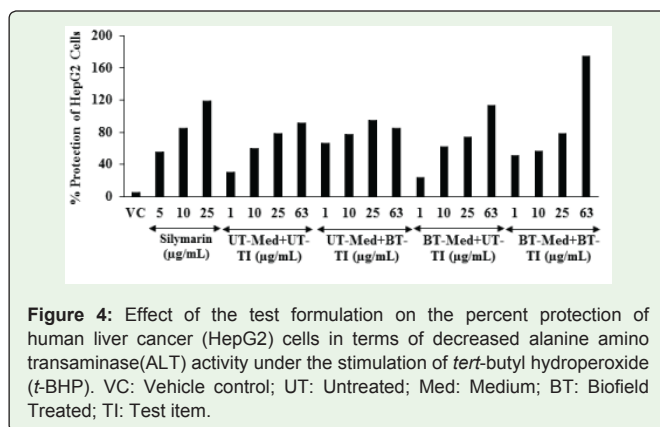
$$\% \text{ Protection} = \frac{[(\text{Absorbance}_{\text{sample}} - \text{Absorbance}_{\text{t-BHP}})] * 100}{[\text{Absorbance}_{\text{untreated}} - \text{Absorbance}_{\text{t-BHP}}]} \tag{2}$$

**Assessment of alkaline phosphatase (ALP) activity**

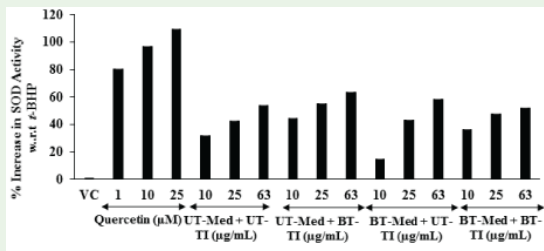
The cells (human bone osteosarcoma cells-MG-63 and human endometrial adenocarcinoma cells-Ishikawa) were counted using a



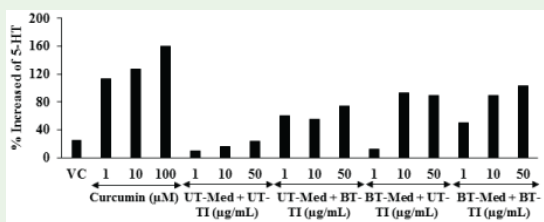
**Figure 3:** The effect of the test formulation on the percent protection of HCF cells in terms of decreased lactate dehydrogenase (LDH) activity against *tert*-butyl hydroperoxide (*t*-BHP) induced damage. TMZ: Trimetazidine; VC: Vehicle control; UT: Untreated; Med: Medium; BT: Biofield Treated; TI: Test item.



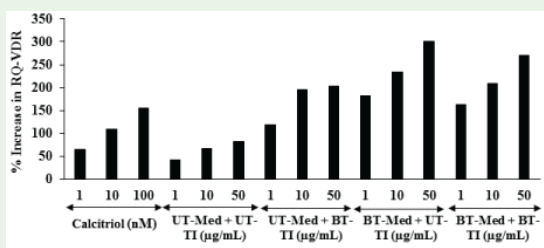
**Figure 4:** Effect of the test formulation on the percent protection of human liver cancer (HepG2) cells in terms of decreased alanine amino transaminase (ALT) activity under the stimulation of *tert*-butyl hydroperoxide (*t*-BHP). VC: Vehicle control; UT: Untreated; Med: Medium; BT: Biofield Treated; TI: Test item.



**Figure 5:** Effect of the test formulation on the percent protection of lungs cells (A549) in terms of increased SOD activity under the stimulation of *tert*-butyl hydroperoxide (*t*-BHP). VC: Vehicle control; UT: Untreated; Med: Medium; BT: Biofield Treated; TI: Test item.



**Figure 6:** Effect of the test formulation on percent increase in 5-hydroxy tryptamine (5-HT) or serotonin in human neuroblastoma cells (SH-SY5Y). VC: Vehicle control; UT: Untreated; Med: Medium; BT: Biofield Treated; TI: Test item.



**Figure 7:** Effect of the test formulation on percent increase in relative quantification (RQ) of vitamin D receptors (VDRs) gene in human bone osteosarcoma cells (MG-63). UT: Untreated; Med: Medium; BT: Biofield Treated; TI: Test item.

hemocytometer and plated in 24-well plates at the density corresponding to  $1 \times 10^4$  cells/well in phenol-free DMEM supplemented with 10% CD-FBS. Following the respective treatments, the cells in the above plate were incubated for 48 hours in  $CO_2$  incubator at  $37^\circ C$ , 5%  $CO_2$ , and 95% humidity. After 48 hours of incubation, the plates were taken out and processed for the measurement of ALP enzyme activity. The cells were washed with  $1 \times PBS$  and lysed by freeze-thaw method i.e., incubation at  $-80^\circ C$  for 20 minutes followed by incubation at  $37^\circ C$  for 10 minutes. To the lysed cells,  $50 \mu L$  of substrate solution i.e., 5 mM of *p*-nitrophenyl phosphate (*p*NPP) in 1M diethanolamine and 0.24 mM magnesium chloride ( $MgCl_2$ ) solution (pH 10.4) was added to all the wells followed by incubation for 1 hour at  $37^\circ C$ . The absorbance of the above solution was read at 405 nm using Synergy HT microplate reader (Biotek, USA). The absorbance values obtained were normalized with substrate blank (*p*NPP solution alone) absorbance values. The percentage increase in ALP enzyme activity with respect to the untreated cells (baseline group) was calculated using Equation 3:

$$\% \text{ Increase in ALP} = \{(X-R)/R\} \times 100 \tag{3}$$

Where, X = Absorbance of cells corresponding to positive control and test groups

R = Absorbance of cells corresponding to baseline group (untreated cells)

**Estimation of lactate dehydrogenase (LDH) in human cardiac fibroblasts (HCF)**

The human cardiac fibroblasts (HCF) Cells were counted and plated at the density of  $0.25 \times 10^6$  cells/ well in 24-well plates in cardiac fibroblast specific medium followed by overnight incubation. The cells were then treated with the test formulation/positive control at the non-cytotoxic concentrations for 24 hours. After 24 hours, oxidative stress was given to the cells using 10mM *t*-BHP for 3.5 hours. The untreated cells were served as control that did not receive any treatment and were maintained in cell growth medium only. Cells treated with 10mM of *t*-BHP alone served as the negative control. After 3.5 hours of incubation with *t*-BHP the above plates were taken out and LDH activity was determined using LDH activity kit as per manufacturer’s instructions. The percent increase in LDH activity was calculated using Equation 4.

$$\% \text{ Increase} = \frac{[(LDH \text{ activity}_{\text{sample}} - LDH \text{ activity}_{t-BHP})] \times 100}{[LDH \text{ activity}_{\text{untreated}} - LDH \text{ activity}_{t-BHP}]} \tag{4}$$

**Estimation of ALT in liver cells (HepG2)**

The human hepatoma cells (HepG2) were counted and plated at the density of  $5 \times 10^4$  cells/well in 48-well plates in DMEM media followed by overnight incubation. The cells were then treated with the test formulation/positive control at the non-cytotoxic concentrations for 24 hours. After 24 hours, oxidative stress was given to the cells using  $400 \mu M$  *t*-BHP for 3.5 hours. The untreated cells served as control that did not receive any treatment and were maintained in cell growth medium only. Cells treated with  $400 \mu M$  of *t*-BHP alone served as negative control. After 3.5 hours of incubation with *t*-BHP the above plates were taken out and ALT activity was determined using ALT activity kit as per manufacturer’s instructions. The percent increase in ALT activity was calculated using Equation 5.

$$\% \text{ Increase} = \frac{[(ALT \text{ activity}_{\text{sample}} - ALT \text{ activity}_{t-BHP})] \times 100}{[ALT \text{ activity}_{\text{untreated}} - ALT \text{ activity}_{t-BHP}]} \tag{5}$$

**Estimation of superoxide dismutase (SOD) in lung (A549) cells**

The adenocarcinomic human alveolar basal epithelial cells (A549) were counted and plated at the density of  $1 \times 10^4$  cells/well in 24-well plates in DMEM followed by overnight incubation. The cells were then treated with the test formulation/ positive control at the non-cytotoxic concentrations along with  $100 \mu M$  *t*-BHP to induce oxidative stress. The untreated cells served as control that did not receive any treatment and were maintained in cell growth medium only. Cells treated with  $100 \mu M$  of *t*-BHP alone served as negative control. After 24 hours of incubation with *t*-BHP the above plates were taken out and SOD activity was determined using SOD activity kit as per manufacturer’s instructions. The percent increase in SOD activity was calculated using Equation 6:

$$\% \text{ Increase in SOD activity} = \{(X-R)/R\} \times 100 \tag{6}$$

Where, X = SOD activity corresponding to Test Item or Positive Control

R = SOD activity corresponding to Control group.

#### Estimation of serotonin in neuronal cells (SH-SY5Y)

The human neuroblastoma (SH-SY5Y) cells were counted and plated at the density of  $10 \times 10^4$  cells/well in 96-well plates followed by overnight incubation. The cells were then treated with the test items/positive control at the non-cytotoxic concentrations. The untreated cells served as control that did not receive any treatment and were maintained in cell growth medium only. The treated cells were incubated for 24 hours. Serotonin release was determined by ELISA as per manufacturer's protocol. The percent increase in serotonin levels was calculated using Equation 7.

$$[(X-R)/R]*100 \quad (7)$$

Where, X = Serotonin levels corresponding to test item or positive control

R = Serotonin levels corresponding to control group.

#### Effect of test formulation on vitamin D receptor (VDR) in bone (MG-63) cells

The human bone osteosarcoma (MG-63) cells were counted using the hemocytometer were plated at a density of  $2 \times 10^5$  cells/well in 6-well plates followed by overnight incubation. The cells were then sera starved for 24 hours and treated with the test formulation/positive control at the non-cytotoxic concentrations. The untreated cells that served as control that did not receive any treatment and were maintained in cell growth medium only. The treated cells were incubated for 24 hours and VDR expression was determined by Q-PCR using VDR specific primers. Cells were harvested by scrapping and washed with PBS. Cell pellets obtained were analyzed for VDR gene expression using human VDR specific primers: Forward: 5'-GCTGACCTGGTCAGTTACAGCA-3', Reverse: 5'-CACGTCACCTGACGCGTACTT-3'. VDR gene expression was normalized using House-keeping (HK) reference. Relative quantification (RQ) of VDR gene in Biofield Energy Treated cells was calculated with respect to the untreated cells using Equation 8:

$$RQ = 2^{-N} \quad (8)$$

Where N is the relative Threshold Cycle (CT) value of treated sample with respect to the untreated sample.

#### Statistical analysis

All the values were represented as percentage. The statistical analysis was performed using SigmaPlot statistical software (v11.0). For two groups comparison student's *t*-test was used. For multiple group comparison, one-way analysis of variance (ANOVA) was used followed by post-hoc analysis by Dunnett's test. Statistically significant values were set at the level of  $p \leq 0.05$ .

## Results and Discussion

### Cell viability using MTT assay

Determination of non-cytotoxic concentration of the formulation and positive controls by MTT cell viability assay was used in terms of

percent viable cells in six (6) different cell-lines *viz.* MG-63, Ishikawa, A549, HepG2, HCF, and SH-SY5Y. Based on the percent cell viability data, it was observed that the formulation and positive controls were safe and non-toxic at the tested concentrations in six different cell lines and selected for other parameters analysis.

### Evaluation of cytoprotective effect of the test formulation

For the assessment of vital organ protection and/or function for the heart, liver, and lungs of the formulation was examined in the *in vitro* cell-based assays under the stimulation of *tert*-butyl hydroperoxide (*t*-BHP) induced oxidative stress. *t*-BHP has been extensively used for the induction of oxidative stress in various cell-lines [41, 42]. The cytoprotective activity of the Biofield Energy Treated test formulation on the restoration of cell viability was determined against *t*-BHP induced cell damage and the result is shown in Figure 1. Trimetazidine (TMZ) was used as positive control in human cardiac fibroblasts cells (HCF) and showed, restoration of cell viability by 94.09% and 102.29% at 10 and 50  $\mu\text{g/mL}$ , respectively compared to the *t*-BHP induced group. Besides, the test formulation showed 112.6%, 44.4%, and 108.65% restoration of cell viability at 1  $\mu\text{g/mL}$  in the UT-Med + BT-TI, BT-Med + UT-TI, BT-Med + BT-TI groups, respectively as compared to the UT-Med + UT-TI group. Moreover, at 10  $\mu\text{g/mL}$  the BT-Med + BT-TI group showed 14.47% restoration of cell viability than UT-Med + UT-TI group (Figure 1). Silymarin was used as positive control in human hepatoma cells (HepG2) resulted, restoration of cell viability by 38.79%, 73.92%, and 81.74% at 5, 10 and 25  $\mu\text{g/mL}$ , respectively compared to the *t*-BHP induced group. Besides, the test formulation showed 845.63% and 60.81% restoration of cell viability at 1  $\mu\text{g/mL}$  in the UT-Med + BT-TI and BT-Med + UT-TI groups, respectively as compared to the UT-Med + UT-TI group. Moreover, at 10  $\mu\text{g/mL}$  the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups showed 85.74%, 32.88%, and 31.54% restoration of cell viability, respectively than UT-Med + UT-TI group. Further, 50.68% and 75.71% cells were restored by UT-Med + BT-TI and BT-Med + UT-TI groups, respectively at 25  $\mu\text{g/mL}$  as compared to UT-Med + UT-TI group (Figure 1). Quercetin was used as positive control in adenocarcinomic human alveolar basal epithelial cells (A549) resulted, restoration of cell viability by 31.24%, 41.93%, and 55.74% at 1, 10 and 25  $\mu\text{g/mL}$ , respectively compared to the *t*-BHP induced group. Besides, the test formulation showed 131.86% restoration of cell viability at 1  $\mu\text{g/mL}$  in the UT-Med + BT-TI group compared to the UT-Med + UT-TI group. Moreover, at 25  $\mu\text{g/mL}$  the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups showed 15.13%, 24.88%, and 27.12% restoration of cell viability, respectively than UT-Med + UT-TI group (Figure 1). The cellular antioxidant capacity can reduce due to excess production of free radicals that leads to inflammation [43]. This excess level of free radicals can affect the normal functions of cell membrane, and ultimately altered the genetic materials and cause various age-related disorders such as diabetes, cardiovascular, autoimmune diseases, and cancer [44-46]. The results suggest that Biofield Treatment has significantly protects *t*-BHP induced cardiotoxicity, hepatotoxicity, and lung cell toxicity which could be due to The Trivedi Effect®. Therefore, Biofield Energy Healing Treatment could be used for the management of cardiovascular, liver, and various lung disorders.

### Assessment of Alkaline Phosphatase (ALP) Activity

The ALP activity was significantly increased by 3.95% and 41.86% in the UT-Med + BT-TI and BT-Med + BT-TI groups, respectively compared to the UT-Med + UT-TI group at 0.1 µg/mL. The effect of the test formulation on bone-specific alkaline phosphatase level is shown in Figure 2. The positive control, calcitriol showed 33.70%, 62.95%, and 76.27% increased the level of ALP at 1, 10, and 100 nM, respectively in MG-63 cells. Moreover, the experimental groups showed 41.86% increased the level of ALP in the BT-Med + BT-TI group with respect to the UT-Med + UT-TI group at 1 µg/mL. At 10 µg/mL, the percent ALP was significantly increased by 94.87%, 99.06%, and 105.13% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively compared to the UT-Med + UT-TI group (Figure 2). Besides, the positive control naringenin showed 39.43%, 88.45%, and 113.64% increased the level of ALP at 1, 10, and 100 nM, respectively in Ishikawa cells. ALP percent was significantly increased by 150.97%, 382.08%, and 471.4% in the BT-Med + UT-TI and BT-Med + BT-TI groups, respectively compared to the UT-Med + UT-TI group at 0.1 µg/mL. Moreover, the experimental groups showed 135.78%, 274.88%, and 364.43% increased the level of ALP in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively with respect to the UT-Med + UT-TI group at 1 µg/mL. At 10 µg/mL, the percent ALP was significantly increased by 40%, 94.44%, and 133.33% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively compared to the UT-Med + UT-TI group (Figure 2). The ALP activity is essential for the bone mineralization and considered a useful biochemical marker for bone formation [47]. Thus, for the detection of bone specific biochemical marker in serum can be clinically useful in evaluating the progress of the bone healing process [48,49]. In this experiment, it was revealed that the Biofield Energy Healing Treated test formulation significantly increased the level of ALP expression, which might be very helpful to the patients suffering from various bone-related disorders.

### Estimation of lactate dehydrogenase (LDH) activity in human cardiac fibroblasts (HCF)

The distribution of lactate dehydrogenase (LDH) is mainly abundant in the heart and skeletal muscle, is a tetrameric enzyme, and is mainly responsible for anaerobic respiration of cells [50-52]. The effect of test items on the percent protection of HCF cells in terms of decreased level of lactate dehydrogenase (LDH) activity is presented in Figure 3. The positive control, trimetazidine (TMZ) exhibited 30.14%, 69.42%, and 80.06% protection of HCF cells (decreased of LDH activity) compared to the *t*-BHP group. The percent protection of HCF cells (decreased of LDH activity) was significantly increased by 35.16%, 115.1%, and 52.32% at 1 µg/mL in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively as compared to the UT-Med + UT-TI group. Moreover, at 10 µg/mL, the percent protection of HCF cells (decreased of LDH activity) was significantly increased by 53.42%, 53.42%, and 165.77% in the UT-Med + BT-TI, BT-Med + UT-TI and BT-Med + BT-TI groups, respectively as compared to the UT-Med + UT-TI group. Further, percent protection of HCF cells (decreased of LDH activity) was also significantly increased by 76.21%, 91.35%, and 85.73% in the UT-Med + BT-TI, BT-Med + UT-TI and BT-Med + BT-TI groups, respectively at 25 µg/mL as compared to the UT-Med + UT-TI group (Figure 3).

Overall, data found that there was a significant reduction of LDH level after Biofield Energy Treatment and protect HCF cells, which might be helpful to resist against various pathological conditions like tissue injury, necrosis, hypoxia, hemolysis or malignancies. It also indicating that the heart cells acted normally under stress and anaerobic condition and improved overall heart function.

### Estimation of alanine amino transferase (ALT) activity in HepG2 cells

The effect of the test formulation on protection of HepG2 cells in terms of decrease alanine amino transferase (ALT) activity is shown in Figure 4. The positive control, silymarin exhibited 56%, 84.98%, and 118.94% protection of HepG2 cells (decreased of ALT activity). The protection of HepG2 cells (decreased of ALT activity) was significantly increased by 117.65% and 67.26% at 1 µg/mL in the UT-Med + BT-TI and BT-Med + BT-TI groups, respectively as compared to the UT-Med + UT-TI group. Moreover, at 10 µg/mL, percent protection of HepG2 cells (decreased of ALT activity) was increased by 30.33% and 4.45% in the UT-Med + BT-TI and BT-Med + UT-TI groups, respectively as compared to the UT-Med + UT-TI group. Further, protection of HepG2 cells (decreased of ALT activity) was also significantly increased by 21.2% in the UT-Med + BT-TI group at 25 µg/mL as compared to the UT-Med + UT-TI group. Further, the percent protection of HepG2 cells (decreased of ALT activity) was increased by 24.4% and 91.3% in the BT-Med + UT-TI and BT-Med + BT-TI groups, respectively at 63 µg/mL as compared to the UT-Med + UT-TI group (Figure 4). The aminotransferase enzymes catalyze the reversible transformation of  $\alpha$ -ketoacids into amino acids. Increased level of ALT is directly proportional to the severity of the diseases like hepatocellular injury and death [53]. Thus, the elevation of ALT enzyme chances of liver disorders [54]. Here, the Biofield Energy Treatment significantly protect liver hepatocytes in terms of reducing the level of transaminases enzyme, ALT compared to the *t*-BHP inducing group.

### Estimation of superoxide dismutase (SOD) activity in adenocarcinomic human alveolar basal epithelial cells (A549)

The effect of the test formulation on the protection of lungs cells (A549) in terms of increased super oxide dismutase (SOD) activity is shown in Figure 5. The positive control, showed 80.67%, 97.01%, and 109.56% protection of A549 (lungs) cells (increased of SOD activity) compared to the *t*-BHP group. The percent protection of A549 (lungs) cells (increased of SOD activity) was significantly increased by 40.56% and 15.17% at 10 µg/mL in the UT-Med + BT-TI and BT-Med + BT-TI groups, respectively compared to the UT-Med + UT-TI group. Moreover, at 25 µg/mL, the percent protection of A549 (lungs) cells (increased of SOD activity) was significantly increased by 30.14% and 12.23% in the UT-Med + BT-TI and BT-Med + BT-TI groups, respectively as compared to the UT-Med + UT-TI group. Further, the percent protection of A549 (lungs) cells (increased of SOD activity) was significantly increased by 16.85% and 8.31% in the UT-Med + BT-TI and BT-Med + UT-TI groups, at 63 µg/mL respectively compared to the UT-Med + UT-TI group (Figure 5). SOD enzyme is considered as an important antioxidant defense mechanism in all living cells which are exposed to oxygen. It possesses a powerful anti-inflammatory activity against chronic inflammation such as colitis.

SOD enzyme supplement treatment reduced the ROS generation, oxidative stress and also inhibits the endothelial activation [55]. Overall, data found that there was a significant increased SOD level after Biofield Energy Treatment in A549 cells, which might be helpful to resist against various pathological conditions like oxidative stress and related adverse effect. It also indicating that the lung cells acted normally and improved overall respiratory activities.

#### Effect of test formulation on serotonin in human neuroblastoma (SH-SY5Y) cells

The effect of test formulation on serotonin level was assessed in SH-SY5Y cells after 24 hours of treatment by ELISA and the results are shown in Figure 6. The positive control, showed 349.48%, 406.69%, and 539.29% increased the level of serotonin compared to the vehicle control (VC) group. The level of serotonin was significantly increased by 543.84%, 28.40%, and 438% at 1 µg/mL in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively compared to the UT-Med + UT-TI group. Moreover, at 10 µg/mL, 5-HT level was significantly increased by 243.58%, 477.12%, and 457.22% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively as compared to the UT-Med + UT-TI group. Further, the serotonin level was significantly increased by 219.98%, 291.29%, and 351.59% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively as compared to the UT-Med + UT-TI group (Figure 6). Serotonin (5-HT) is widely distributed in CNS, and its role is highly important in various neuronal functions like sleep, feeding, pain, sexual behavior, cardiac regulation, and cognition. Loss of serotonin leads to various neuropsychiatric diseases of late life such as depression, Alzheimer's disease, loss of ability of thinking, memory loss, cognitive health, etc. [56]. Thus, the data suggested that Biofield Energy Healing Treated novel test formulation significantly improved the serotonin level, which would be highly useful against various neurodegenerative diseases and other age-related disorders and improved the normal functioning of the brain tissues.

#### Effect of test formulation on vitamin D receptors (VDRs)

Human bone osteosarcoma cells (MG-63) were treated with the test formulation and the effect on VDR expression was determined using quantitative-polymerase chain reaction (Q-PCR) amplification. VDR-relative threshold cycle (VDR-CT) values were obtained from PCR amplification. Relative quantification (RQ) was calculated from the VDR-CT and house-keeping (HK)-CT values for MG-63 cells treated with test formulation and positive control is represented in Figure 7. The RQ of VDR was increased in a concentration-dependent manner by 65.86%, 109.94%, and 154.91% in positive control group (calcitriol) at 1, 10, and 100 nM, respectively. Moreover, RQ of VDR was significantly increased by 186.96%, 341.43%, and 291.31% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively at 1 µg/mL compared to the UT-Med + UT-TI group. Additionally, at 10 µg/mL the VDR level was significantly increased by 190.73%, 249.21%, and 212.61% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively compared to the UT-Med + UT-TI group. Further, VDR level was also significantly increased by 149.25%, 268.1%, and 232.07% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively at 50 µg/mL compared to the UT-Med + UT-TI group. The most abundant

vitamin D metabolite is 25-hydroxyvitamin D3 (25(OH)D3), has been used as a biomarker for the vitamin D status of the human body. However, the biologically most active vitamin D compound is 1α, 25-dihydroxyvitamin D3 (1, 25(OH)2D3), which functions as specific high-affinity ligand of the transcription factor of VDRs [57,58]. Overall, the Biofield Energy Treated test formulation has significantly increased the expression of VDRs, which might be helpful to bind more active vitamin D3 metabolites and that ultimately can improve the more physiological functions of vitamin D and simultaneously improved bone cell growth and development.

#### Conclusions

The study outcomes showed that the tested novel Biofield Energy Treated formulation was safe and non-toxic based on MTT cell viability assay in six tested cells. The treatment groups like UT-Med + BT-TI and BT-Med + BT-TI showed 112.6% and 108.65% restoration of cell viability in human cardiac fibroblasts cells (HCF) compared to UT-Med + UT-TI group. Moreover, UT-Med + BT-TI group showed 845.63% restoration of cell viability at 1 µg/mL in human hepatoma cells (HepG2) compared to untreated. Alkaline phosphatase (ALP) activity was significantly increased by 150.97%, 382.08%, and 471.4% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively at 0.1 µg/mL in human endometrial adenocarcinoma cells (Ishikawa). The percent protection of HCF cells (decreased of LDH activity) was significantly increased by 76.21% (at 25 µg/mL), 115.1% (at 1 µg/mL), and 165.77% (at µg/mL) in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively compared to the untreated in HCF cells. The percent protection of HepG2 cells (decreased of ALT activity) was significantly increased by 117.65% (at 1 µg/mL) and 91.3% (at 63 µg/mL) in the UT-Med + BT-TI and BT-Med + BT-TI groups, respectively compared to untreated group in HepG2 cells. The percent protection of A549 (lungs) cells (increased of SOD activity) was significantly increased by 40.56%, 30.14%, and 16.85% in the UT-Med + BT-TI group at 10, 25, and 63 µg/mL, respectively compared to untreated in A549 cells. The serotonin level was significantly increased by 543.84% and 438% at 1 µg/mL in the UT-Med + BT-TI and BT-Med + BT-TI groups, respectively compared to the untreated in human neuroblastoma cells (SH-SY5Y). The relative quantification (RQ) of vitamin D receptors (VDRs) level was significantly increased by 186.96%, 341.43%, and 291.31% in the UT-Med + BT-TI, BT-Med + UT-TI, and BT-Med + BT-TI groups, respectively at 1 µg/mL compared to the untreated in MG-63 cells. In conclusion, The Biofield Energy Treatment significantly improved heart, liver, bones, neuronal, and lungs functional enzyme biomarkers and also protected cardiomyocyte, hepatocyte, osteocytes, pneumocyte, and nerve cells from oxidative damage induced by *tert*-butyl hydroperoxide (*t*-BHP). Thus, results suggested that Biofield Energy Treatment can be used as a complementary and alternative treatment for the prevention of various types of cardiac disorders (high blood pressure, stroke, congestive heart failure, peripheral artery disease, congenital heart disease, rheumatic heart disease, valvular heart disease, carditis, thromboembolic disease, and venous thrombosis, etc.), hepatic disorders (cirrhosis, liver cancer, hemochromatosis, Wilson disease), and lungs disorders (Asthma, Chronic bronchitis, Emphysema, Cystic fibrosis, Pneumonia). Further, it can be useful to improve cell-to-cell

messaging, normal cell growth and differentiation, cell cycling and proliferation, neurotransmission, skin health, hormonal balance, immune and cardiovascular functions. Moreover, it can also be utilized in organ transplants (i.e., kidney, liver, and heart transplants), hormonal imbalance, aging, and various inflammatory and immune-related disease conditions like Alzheimer's Disease (AD), Ulcerative Colitis (UC), Dermatitis, Asthma, Irritable Bowel Syndrome (IBS), Hashimoto Thyroiditis, Pernicious Anemia, Sjogren Syndrome, Multiple Sclerosis, Aplastic Anemia, Hepatitis, Graves' Disease, Dermatomyositis, Diabetes, Parkinson's Disease, Myasthenia Gravis, Atherosclerosis, Systemic Lupus Erythematosus (SLE), stress, etc. to improve overall health and Quality of Life.

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